NEW KAURANE DITERPENOIDS FROM THE ROOTS OF ELAEOSELINUM TENUIFOLIUM¹

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ABSTRACT.—The C₆H₆ extract from the roots of *Elaeoselinum tenuifolium* afforded two new tetracyclic diterpenes identified as *ent*-15 α -angeloyloxykaur-16-en-3 β -ol and *ent*-15 α -angeloyloxy-16 β , 17-epoxykauran-3 β -ol, as evidenced by spectral data and chemical transformations. 2-Isopropylmethylanisole, thymoquinol dimethyl ether, apiol, and β -sitosterol were also isolated.

In our search for new natural substances from plants endemic to Comunidad Valenciana (East Spain), we have examined the chemical constituents of the roots of *Elaeoselinum tenuifolium* (Lag) Lange (Umbelliferae), also known as *Thapsia tenuifolia* Lag. and *Elaeoselinum lagascae* Boiss.

The most characteristic components found in the Umbelliferae are coumarins, terpenoids, and aromatic derivatives from the essential oils, as well as phenylpropanoids and flavonoids, usually minor components (1). The diterpenes are quite uncommon in these plants; however, some of them have been reported: labdane acids from *Hermes villosa* (2), magydardienediol and other monocyclic derivatives from *Magydaris panacifolia* (3), and also tetracyclic diterpenes from *Elaeoselinum gummiferum* (4-6) and *Elaesoselinum foetidum* (7). We have also found diterpenes with beyerane, kaurane, and atisane skeletons in the roots of *Elaesoselinum asclepium* (8), and in the present study, we report the structural identification of the new kaurane derivatives **1** and **14**.

RESULTS AND DISCUSSION

The C_6H_6 extract from the roots of the plant (3.9%, dry weight) was defatted with MeOH, and the soluble portion was separated into neutral and acidic fractions with 4% aqueous NaOH. The neutral fraction in Et₂O afforded, on standing, a crystalline product from which compounds **1** and **14** were isolated by chromatography on silica gel. The noncrystalline material provided further amounts of the above compounds and sitosterol (9), 2-isopropyl-4-methylanisole (10), thymoquinol dimethyl ether (11), and apiol (12).

Compound **1**, which represents nearly one-third of the C₆H₆ extract, showed in the ms a molecular ion M⁺ at m/z 386 in agreement with the formula C₂₅H₃₈O₃. The ir spectrum showed absorption bands of hydroxyl (3360 cm⁻¹), conjugated ester (1715 cm⁻¹) and methylidene groups (3070, 1650, 880 cm⁻¹). The ¹H-nmr spectrum confirmed the presence of an exocyclic methylene group [δ 4.91 (d, J=2.4 Hz, H-17b), 4.87 (m, W¹/₂=5 Hz, H-17a)], one proton geminal to an ester group [δ 5.23 (t, J=2.4 Hz, H-15)], and one proton geminal to a hydroxyl group [δ 3.17, m, X (ABX), J_{AX+BX} =16.3 Hz, H-3].

The ester group was identified as an angelate as deduced from the characteristic ¹Hnmr signals [δ 6.07 (qq, J=7.2 and 1.4 Hz, H-3'), 2.00 (dq, J=7.2 and 1.4 Hz, Me-

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4') and 1.91 (qnt, J=1.4 Hz, Me-5')], ¹³C-nmr signals [δ 168.0 (s, C-1'), 138.1 (d, C-3'), 128.1 (s, C-2'), 20.8 (q, Me-5'), 15.8 (q, Me-4')] (5, 13), as well as from the ms fragments at m/z 286, 83, and 55 (14). Furthermore, the alkaline hydrolysis of **1** with 5% KOH/MeOH yielded tiglic acid, isomerization product of angelic acid, and the alcohol **3**.

According to the molecular formula, the ¹H- and the ¹³C-nmr data, it was concluded that **1** was a tetracyclic diterpene with one methylidene group, one equatorial secondary hydroxyl group, and one secondary angeloyloxy group. These functional groups can be accommodated on the common tetracyclic diterpene skeletons beyerane, atisane, kaurane, or phyllocladane. The presence of one exocyclic methylene and three quaternary methyl groups in **1** (δ 1.02, 0.93, 0.74) let us discount a beyerane skeleton. The chemical shift of the methinic allylic proton (H-12 in atisane, H-13 in kaurane and phyllocladane), which resonates at δ 2.66, also allowed us to discard the atisane skeleton. In atisane derivatives, the allylic proton H-12 generally absorbs at ca. δ 2.3 ppm (6).

The ¹³C-nmr spectrum supported a kaurane skeleton for **1** and allowed us to assign the position of the substituents. According to the available data (5, 6, 15-19), the chemical shift for the quaternary C-10 in kauranes appears at δ >38.8, perhaps because of a γ -gauche effect between C-10 and C-12, whereas for phyllocladane derivatives (C-10 and C-12 *anti*), the C-10 signal occurs at higher fields (δ <37.8). As all carbon singlet signals of **1** appear above δ 38.8 (see Experimental section), we tentatively assigned a kaurane skeleton to our diterpene.

The location of the angeloyloxy and hydroxy groups in the 16-kaurene skeleton was deduced as follows. The signal of the geminal proton to the angeloyloxy group [δ 5.23 (t, J=2.4 Hz)] collapses to a singlet by irradiation of the exocyclic methylidene protons, and this fixed the ester group at C-15. The signal of the geminal proton to the free hydroxyl appears as a multiplet at 3.17 (six lines, $J_{AX+BX}=16.3$ Hz) (20), also observed in other terpenoids when an equatorial hydroxyl group is flanked by a quaternary carbon and a -CH₂- group, and, consequently, the secondary hydroxyl group could be placed either at C-1, at C-3, or at C-7. This hydroxyl group was placed at C-3 according to the ¹³C-nmr data, fully consistent with the chemical shifts observed for other 3 β -hydroxy polycyclic diterpenoids and triterpenoids (19).

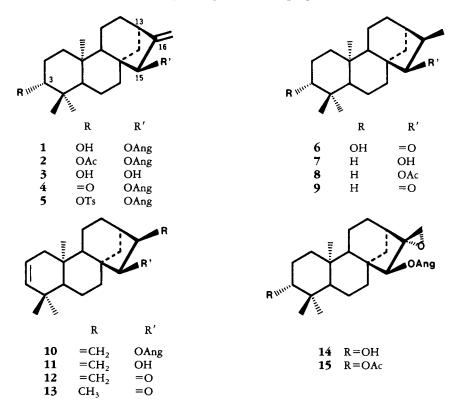
The observed deshielding for H-15 in compounds **1-5** suggests the *endo*-(=*ent*-15 α) configuration for the angeloyloxy group. Also the ¹³C-chemical shifts calculated for the *endo*-configuration of the 15-hydroxyl group (16) are nearly the same as observed ($\Delta\delta$ <0.4 ppm), but the calculated shifts for the *exo*-15-hydroxyl are rather different from those observed, particularly for C-8, C-9, C-13, C-14, C-16, and C-17 ($\Delta\delta$ 2-3 ppm). The *endo*-configuration was also supported by chemical transformation of diol **3**. Treatment of this diol with concentrated HCl/MeOH easily gave the rearranged ketone **6**. This rearrangement is difficult in the case of the *exo*-(=*ent*-15 β) configuration (21).

The absolute configuration for **1** was proposed according to the cd spectrum of **4**. The observed Cotton effect [$\Delta \epsilon - 0.97$ (293 nm)] suggests that these kaurane derivatives belong to *enantio*-series.

Lastly, the structure of 1 was confirmed by transformation into the known kaurane derivatives 7 and 9 (22). Kauranol 7 was first isolated in low yield from the Huang-Minlon reduction products from 4. Further amounts of 7 were obtained as follows. Dehydration of 1 via tosylate 5 gave the alkene 10 which was hydrolyzed to $11.^2$ Catalytic hydrogenation of 11 (PtO₂/HOAc) yielded a mixture of 7 and 9. Rearrangement of 11

with concentrated HCl/MeOH to give 13, followed by catalytic hydrogenation, yielded only the ketone 9.

The natural compound 14 showed a molecular ion at m/z 402, which corresponds to the molecular formula $C_{25}H_{38}O_4$. The spectra of 1 and 14 are quite similar, but the ¹H-nmr and ir traces of 14 revealed the absence of the exocyclic methylidene group signals. On the contrary, the nmr spectrum of 14 showed a broad singlet at δ 2.80 which was assigned to an epoxidic methylene group. The proposed structure for 14 was confirmed by epoxidation of 1 with *m*-CPBA, which results in a compound identical in all aspects to 14. Assuming that the epoxidation took place by peroxyacid attack from the less hindered *exo*-side, the *ent*-16 β configuration was proposed for 14.



EXPERIMENTAL

GENERAL EXPERIMENTAL PROCEDURES. —Melting points were determined on a Kofler hot-stage apparatus and are uncorrected. Spectra were recorded with the following instruments: uv, B & L Spectronic 2000; ir, Pye Unicam SP-200 and SP-2000; nmr, Brucker WP 200 (1 H, 200 MHz; 13 C, 50.3 MHz) and Varian EM 360L (60 MHz) recorded in CDCl₃ with TMS as internal standard (scale δ in ppm); e.i., mass spectra, Hewlett-Packard 5930A with direct inlet probe at 70 eV; Optical Activity AA-100 polarimeter; cd spectra, Jobin Yvon Dichrograph Mark III.

EXTRACTION AND ISOLATION.—The plant was collected at "Cabo las Huertas" Alicante, Spain, and a voucher specimen was deposited in the Department of Biology, University of Alicante. The air-dried roots of *E. tenuifolium* (1.34 kg) were extracted with C_6H_6 in a Dean-Stark apparatus. The C_6H_6 extract (58.8 g, 3.9% weight of dried roots) was treated with MeOH, and the soluble portion (45.5 g) was separated with Et_2O and 4% NaOH into neutral and acidic fractions.

The neutral fraction (35.9 g) in Et₂O afforded a crystalline mixture (15.8 g) containing mainly 1 and 14, which were isolated by dry column chromatography with hexane-EtOAc (8:2). The mother liquor (18.8 g) was chromatographed on silica gel (Merck ref. 7733, 500 g) in a column packed with hexane, using hexane/EtOAc mixtures as eluent. The concentration of EtOAc was gradually increased, and 88 fractions, each 250 ml, were collected. Fractions 13-14, eluted with hexane-EtOAc (97:3), contained 2-iso-

propyl-4-methylanisole. Thymoquinol dimethyl ether was isolated by rechromatography from fraction 15, and apiol was isolated from fraction 19. Fractions 53-59, eluted with hexane-EtOAc (9:1), afforded a mixture of two substances. Compound 1 crystallized from the mixture (2.3 g), and sitosterol was purified from the mother liquor after chromatography. Fraction 64-83 eluted with hexane-EtOAc (7:3) afforded 14 (0.6 g), which was purified by crystallization.

ent-15α-Angeloyloxykaur-16-en-3β-ol (1).—Rf 0.4 (hexane-EtOAc, 8:2), needles mp 120-121° (hexane); [α]D = 89° (CHCl₃, c 1.1); uv λ max (EtOH) 218 nm (log ϵ 3.9); ir ν max (KBr) 3360, 3070, 2920, 2860, 1715, 1650, 1450, 1380, 1220, 1150, 1040, 880, 840 cm⁻¹; ¹H nmr (200 MHz) δ 0.74 (3H, s, H-19), 0.93 (3H, s, H-18), 1.02 (3H, s, H-20), 1.91 (3H, qnt, J=1.4 Hz, H-5'), 2.00 (3H, dq, J=7.2 and 1.4 Hz, H-4'), 2.66 (1H, m, H-13), 3.17 (1H, m, X (ABX), J_{AX+BX} =16.3 Hz, H-3), ³ 4.87 (1H, m, W¹/₂=5 Hz, H-17a), 4.91 (1H, d, J=2.4 Hz, H-1b), 5.23 (1H, t, J=2.4 Hz, H-15), 6.07 (1H, qq, J=7.2 and 1.4 Hz, H-3'); ¹³C nmr⁴ δ 168.1 (s, C-1'), 153.9 (s, C-16), 138.1 (s, C-2'), 106.3 (t, C-17), 81.2 (d, C-15), 78.9 (d, C-3), 54.9 (d, C-5), 48.2 (d, C-9), 46.0 (s, C-8), 40.8 (d, C-13), 39.0 (s, C-10*), 39.0 (t, C-14**), 38.8 (s, C-4*), 38.8 (t, C-1**), 36.5 (t, C-7), 33.5 (t, C-12), 28.4 (q, C-18), 27.4 (t, C-2), 20.8 (q, C-5'), 19.6 (t, C-6), 17.8 (q, C-20), 15.8 (q, C-4'), 15.6 (q, C-19); ms m/z (%) 386 [M]⁺ (3), 303 [M-Ang]⁺ (0.5), 286 [M-AngOH]⁺ (20), 271 [286-Me]⁺ (10), 268 [286-H₂O]⁺ (5), 253 [271-H₂O]⁺ (9), 243 [271-C₂H₄]⁺ (7), 225(4), 145(5), 119(7), 105(12), 91(19), 83 [Ang]⁺ (100), 55 [Ang-CO]⁺ (62), 41(47). Anal. calcd for C₂₅H₃₈O₃: C, 77.68; H, 9.91. Found: C, 77.88; H, 10.10.

Acetate 2.—The alcohol 1 (290 mg) in pyridine (1 ml) and Ac₂O (2 ml) was left overnight at room temperature. After usual work up, acetate 2 (275 mg) was isolated, mp 94-95° (hexane); $[\alpha]D - 75°$ (CHCl₃, c 1.2); ir ν max (KBr) 3080, 2940, 2860, 1735, 1715, 1660, 1645, 1480, 1460, 1440, 1380, 1370, 1250, 1150, 885, 760 cm⁻¹; ¹H nmr (60 MHz) δ 0.84 (6H, s, H-18 and H-19), 1.07 (3H, s, H-20), 1.96 (3H, br s, H-5'), 2.03 (3H, br d, J=7 Hz, H-4'), 2.03 (3H, s, H-2"), 2.70 (1H, m, H-13), 4.46 (1H, dd, J=9 and 7 Hz, H-3), 4.92 (2H, br s, H-17), 5.26 (1H, t, J=2.5 Hz, H-15), 6.13 (1H, br q, J=7 Hz, H-3'); ms m/z (%) 428 [M]⁺ (2), 368 [M-AcOH]⁺ (4), 345 [M-Ang]⁺ (2), 328 [M-AngOH]⁺ (5), 313(2), 286(6), 268(6), 253(9), 243(3), 255(3), 145(2), 131(5), 121(6), 105(7), 91(12), 83 [AngOH]⁺ (100), 55 [Ang-CO]⁺ (39), 43(52), 29(13).

ent-*Kaur-16-en-15α*, 3β-*diol* (3).—Compound **1** (185 mg) was refluxed with 5% KOH/MeOH (20 ml) for 7 h. The MeOH was removed and the residue taken up in H₂O and extracted with Et₂O, washed with H₂O, dried and evaporated to yield **3** (135 mg), which was crystallized from C₆H₆ as white prisms; mp 167-168°; [α]D – 61° (CHCl₃, c 1.4); ir ν max (KBr) 3430, 3350, 3060, 2960, 2920, 2830, 1660, 1440, 1280, 1180, 1080, 1060, 1025, 990, 880, 865 cm⁻¹; ¹H nmr δ 0.78 (3H, s, H-19), 0.98 (3H, s, H-18), 1.03 (3H, s, H-20), 2.70 (1H, m, H-13), 3.21 (1H, dd, J=9 and 7 Hz, H-3), 3.75 (1H, t, J=2.4 Hz, H-15), 4.95 (1H, d, J=2.8 Hz, H-17a), 5.09 (1H, m, W¹/₂=5 Hz, H-17b); ¹³C nmr⁴ δ 158.6 (s, C-16), 104.8 (t, C-17), 82.4 (d, C-15), 79.0 (d, C-3), 54.6 (d, C-5), 46.5 (d, C-9), 45.7 (s, C-8), 40.2 (d, C-13), 38.9 (s, C-10*), 38.9 (t, C-14**), 38.8 (t, C-1**), 38.8 (s, C-4*), 36.5 (t, C-7), 33.3 (t, C-12), 28.4 (q, C-18), 27.5 (t, C-2), 19.7 (t, C-6), 18.2 (t, C-11), 17.6 (q, C-20), 15.5 (q, C-19); ms m/z (%) 304 [M]⁺ (35), 289 [M-Me]⁺ (22), 286 [M-H₂O]⁺ (33), 271 [286-Me]⁺ (58), 253 [271-H₂O]⁺ (20), 246(55), 203(28), 173(18), 164(16), 147(33), 121(45), 107(55), 91(63), 84(70), 83(20), 81(57), 67(50), 55(70), 43(57), 41(100).

ent-(16S)-3β-Hydroxykauran-15-one (**6**).—The diol **3** (64 mg) in MeOH (10 ml) and Et₂O (5 ml) was treated with concentrated HCl (2 ml) for 24 h at room temperature. Removal of the solvents and recovery of the product with Et₂O gave **6**; mp 158-159° (hexane); [α]D -80° (CHCl₃, c, 1.0); λ max (EtOH) 204 nm (log ϵ 4.1); cd (hexane) $\Delta\epsilon$ -1.04 (308 nm); cd (MeOH) $\Delta\epsilon$ -0.53 (308 nm) and +0.09 (276 nm); ir ν max (KBr) 3520, 3300, 2920, 2860, 1725, 1470, 1440, 1380, 1365, 1175, 1090, 1036, 990, 920 cm⁻¹; ¹H nmr (200 MH2) δ 0.77 (3H, s, H-19), 0.98 (3H, s, H-18), 1.06 (3H, s, H-20), 1.09 (3H, d, *J*=7 Hz, H-17), 2.22 (1H, qnt, *J*=7 Hz, H-16), 2.42 (1H, m, H-13), 2.43 (1H, br d, *J*=2 Hz, H-14), 3.19 (1H, m, six lines, X (ABX), J_{AX+BX} = 16.3 Hz, H-3); ¹³C nmr δ 202.3 (s, C-15), 78.8 (d, C-3), 54.5 (d, C-5), 52.5 (s, C-8), 52.4 (d, C-9), 47.8 (d, C-16), 39.4 (s, C-10), 38.9 (s, C-4), 37.9 (t, C-1), 37.5 (t, C-7), 35.0 (d, C-13), 34.3 (t, C-14), 28.2 (q, C-18), 27.1 (t, C-2), 24.8 (t, C-12), 18.7 (t, C-6), 18.1 (t, C-11), 17.8 (q, C-20), 15.4 (q, C-19), 10.1 (q, C-17); ms m/z (%) 304 [M]⁺ (65), 289 [M-Me]⁺ (9), 286 [M-H₂O]⁺ (10), 271 [289-H₂O]⁺ (15), 253(8), 246(100), 228(21), 213(43), 135(51), 107(43), 93(45), 79(41), 69(40), 55(50), 41(62).

ent-15 α -Angeloyloxykaur-16-en-3-one (4).—Pyridinium chlorochromate (PCC, 300 mg) was gradually added to a stirred solution of 1 (250 mg) in CHCl₃ (20 ml). After 4 h at room temperature, the mixture was worked up to give 4 (211 mg), gummy; [α]D – 106° (CHCl₃, c 2.3); cd (hexane) $\Delta \epsilon$ -0.97 (293 nm); ir

³This signal at 60 MHz appeared as a dd, J=9 and 7 Hz.

⁴(*, **) Assignments may be reversed.

 ν max (film) 3060, 2930, 2860, 1710, 1450, 1380, 1230, 1150, 1040, 960, 940, 890, 845 cm⁻¹; ¹H nmr δ 1.03 (3H, s, H-19), 1.06 (3H, s, H-18), 1.13 (3H, s, H-20), 1.96 (3H, br s, H-5'), 2.03 (3H, br d, J=7 Hz, H-4'), 2.70 (1H, m, H-13), 4.93 (2H, br s, H-17), 5.30 (1H, t, J=2.5 Hz, H-15), 6.13 (1H, br q, J=7 Hz, H-3'); ms m/z (%) 384 [M]⁺ (1), 369 [M-Me]⁺ (1), 301 [M-Ang]⁺ (1), 284 [M-AngOH]⁺ (5), 269 [284-Me]⁺ (4), 256(1), 227(1), 199(2), 131(3), 93(6), 83 [Ang]⁺ (100), 67(5), 55 [Ang-CO]⁺ (61), 41(23).

ent-(16S)-Kauran-15 α -ol (7).—A solution of **4** (140 mg) in diethyleneglycol (4 ml) was refluxed with 80% hydrazine hydrate (1 ml) for 2 h. KOH (300 mg) was added, refluxed for 45 min and then for 1 h to remove the excess of H₂O and hydrazine hydrate; the remaining solution was refluxed for 3 h. The crude product was extracted with Et₂O and purified on a small silica gel column and crystallized from MeOH; mp 133-134°; $\{\alpha\}D = 50^{\circ}$ (CHCl₃, c 1.8); ir ν max (KBr) 3380, 2980, 2920, 2880, 2850, 1460, 1450, 1365, 1100, 1020, 990 cm⁻¹; ¹H nmr (60 MHz) δ 0.80 (3H, s, H-19), 0.83 (3H, s, H-18), 0.93 (3H, d, J=7 Hz, H-17), 1.01 (3H, s, H-20), 3.60 (1H, d, J=11 Hz, H-15); ms m/z (%) 290 [M]⁺ (58), 275 [M-Me]⁺ (89), 257 [275-H₂O]⁺ (12), 231(29), 137(33), 123(28), 107(34), 91(47), 69(50), 55(70), 41(100), 29(44).

Acetate **8**.—The alcohol **7** (34 mg) upon treatment with Ac₂O/Py as above gave **8** (21 mg); mp 111-112° (MeOH); ir ν max (KBr) 2990, 2940, 2920, 2860, 2840, 1720, 1440, 1365, 1230, 1085, 1050, 1015, 915 cm⁻¹; ¹H nmr (200 MHz) δ 0.80 (3H, s, H-19), 0.83 (3H, d, J=7 Hz, H-17), 0.84 (3H, s, H-18), 1.03 (3H, s, H-20), 2.11 (3H, s, H-2'), 2.36 (1H, ddq, J_{15-16} =11, J_d =7.2 and J_q =7 Hz, H-16), 4.76 (1H, d, J=11 Hz, H-15).

Tosylate (5).—The alcohol **1** (687 mg) was dissolved in pyridine (1C ml) and treated at 0° with *p*-toluensulphonyl chloride (800 mg) for 1 h. The mixture was left at 0° for 48 h and monitored by tlc. The product was extracted with Et_2O , washed with 2 N HCl, 5% NaHCO₃, and H₂O and dried over anhydrous Na₂SO₄. The crude product (503 mg) was crystallized from hexane; mp 130-131°; [α]D -66° (CHCl₃, c 1.9); ir 3080, 2940, 2870, 1700, 1640, 1595, 1440, 1360, 1225, 1175, 1155, 1095, 920, 875, 710, 670 cm⁻¹; ¹H nmr (60 MHz) δ 0.80 (6H, s, H-18 and H-19), 1.03 (3H, s, H-20), 1.96 (3H, br s, H-5'), 2.03 (3H, br d, J=7 Hz, H-4'), 2.42 (3H, s, Me-Ar), 2.67 (1H, m, H-13), 4.26 (1H, dd, J=9 and 7 Hz, H-3), 4.90 (2H, br s, H-17), 5.21 (1H, t, J=2.5 Hz, H-15), 6.13 (1H, br q, J=7 Hz, H-3'), 7.30 (2H, d, J=8.5 Hz, H_m-Ar), 7.80 (2H, d, J=8.5 Hz, H₀-Ar); ms m/z (%) 540 [M]⁺ (5), 497(11), 458(11), 440 [M-AngOH]⁺ (17), 425 [440-Me]⁺ (12), 368(56), 325(35), 286(50), 268(59), 253(55), 225(53), 172(26), 107(29), 83 [Ang]⁺ (100), 55 [Ang-CO]⁺ (46).

ent-15α-Angeloyloxykaur-2, 16-diene (**10**).—A solution of **5** (803 mg) in quinoline was refluxed at 160° for 1.5 h. H₂O was added, and the product was extracted with Et₂O and washed with 2 N HCl, 5% NaHCO₃, and H₂O. Dried over anhydrous Na₂SO₄, the reaction product was purified by column chromatography on silica gel to yield **10** (450 mg), oil; $[\alpha]D - 101^{\circ}$ (CHCl₃, c 1.10); ir ν max (film) 3060, 2920, 2860, 1710, 1645, 1450, 1375, 1360, 1225, 1150, 1045, 990, 965, 940, 885, 845, 750, 735, 720 cm⁻¹; ¹H nmr (60 MHz) δ 0.88 (3H, d, H-19), 0.95 (3H, br d, *J*=7 Hz, H-4'), 2.73 (1H, m, H-13), 4.93 (2H, br s, H-17), 5.26 (1H, t, *J*=2.5 Hz, H-15), 5.40 (2H, d, AB(X), H-2 and H-3), 6.10 (1H, br q, *J*=7 Hz, H-3').

ent-*Kaur-2*, 16-diene-15 α -ol (11).—Compound 10 (350 mg) was hydrolyzed with 5% KOH/MeOH at room temperature for 6 h, affording 11 (219 mg) as an oil; [α]D -69° (CHCl₃, c 2.7); ir ν max (film) 3410, 3060, 2920, 2850, 1655, 1640, 1440, 1370, 1360, 1250, 1120, 1070, 1050, 990, 885, 720 cm⁻¹; ¹H nmr (60 MHz) δ 0.86 (3H, s, H-19), 0.93 (3H, d, H-18), 1.05 (3H, s, H-20), 2.68 (1H, m, H-13), 3.76 (1H, t, *J*=2.5 Hz, H-15), 4.93 (1H, d, *J*=3 Hz, H-17a), 5.06 (1H, m, W¹/₂=5 Hz, H-17b), 5.38 (2H, m, AB(X), H-2 and H-3).

Catalytic hydrogenation of 11 (60 mg) with H_2/PtO_2 -HOAc (see below), afforded a 1:1 mixture of 7 and 9.

ent-*Kaur-2*, 16-diene-15-one (12).—The alcohol 11, on standing in contact with the air at room temperature, was partially oxidized to 12. After chromatography and crystallization from MeOH showed: mp 100-101°; $[\alpha]D = 208^{\circ}$ (CHCl₃, c 1.0); uv λ max (EtOH) 232 nm (log \in 3.8); ir ν max (KBr) 3060, 2920, 2850, 1710, 1630, 1440, 1250, 1190, 1160, 1045, 1035, 960, 940, 925, 715 cm⁻¹; ¹H nmr (60 MHz) δ 0.92 (3H, s, H-19), 0.98 (3H, s, H-18), 1.15 (3H, s, H-20), 3.06 (1H, m, H-13), 5.23 (1H, br s, H-17a), 5.40 (2H, m, AB(X), H-2 and H-3), 5.97 (1H, br s, H-17b); ms m/z (%) 284 [M]⁺ (100), 269 [M-Me]⁺ (32), 251(12), 229(35), 144(22), 136(22), 119(30), 105(41), 91(41), 77(26), 67(14), 55(18), 41(26).

ent-(16S)-Kaur-2-ene-15-one (13).—Treatment of 11 (75 mg) with concentrated HCl in MeOH/ Er₂O (see above 6), afforded 13 (40 mg); mp 107-108° (EtOH); ir ν max (KBr) 3010, 2920, 2860, 1725, 1450, 1370, 725 cm⁻¹; ¹H nmr (60 MHz) δ 0.88 (3H, s, H-19), 0.96 (3H, s, H-18), 1.10 (3H, s, H-20), 1.10 (3H, d, J=7 Hz, H-17), 5.38 (2H, m, AB(X), H-2 and H-3). ent-(16S)-Kauran-15-one (9).—A solution of **13** (40 mg) in HOAc (5 ml) and a catalytic amount of PtO₂ (2 mg) was strongly stirred under H₂ for 6 h at room temperature. After usual work up, **9** was isolated; mp 145-147° (Me₂CO); $[\alpha]D = 92^{\circ}$ (CHCl₃, c 0.3); ir ν max (KBr) 2920, 2860, 1725, 1480, 1450, 1385, 1370, 970, 920 cm⁻¹; ¹H nmr (60 MHz) δ 0.80 (3H, s, H-19), 0.86 (3H, s, H-18), 1.07 (3H, s, H-20), 1.10 (3H, d, J=7 Hz, H-17); ms m/z (%) 288 [M]⁺ (59), 273 [M-Me]⁺ (25), 255(9), 245(13), 230(100), 215(48), 149(12), 137(28), 123(40), 121(15), 107(23), 91(31), 55(36), 41(39).

ent-15 α -Angeloyloxy-16 β , 17-epoxykaurane-3 β -ol (14).—Isolated from fractions 64-83 of the main chromatography. Needles; mp 129-130° (hexane); $[\alpha]D - 22°$ (CHCl₃, c 1.3); ir ν max (KBr) 3360, 2940, 2870, 1715, 1650, 1460, 1380, 1230, 1150, 1045, 850 cm⁻¹; ¹H nmr (60 MHz) δ 0.78 (3H, s, H-19), 0.96 (3H, s, H-18), 1.07 (3H, s, H-20), 1.93 (3H, br s, H-5'), 2.03 (3H, br d, J=7 Hz, H-4'), 2.80 (2H, s, H-17), 3.20 (1H, dd, J=9 and 7 Hz, H-3), 5.03 (1H, br s, H-15), 6.13 (1H, br q, J=7 Hz, H-3'); ms m/z (%) 402[M]⁺ (0.5), 384 [M-H₂O]⁺ (6), 284 [302-H₂O]⁺ (7), 269 [284-Me]⁺ (7), 251(3), 135(9), 91(10), 83 [Ang]⁺ (100), 81(10), 79(9), 69(9), 55 [Ang-CO]⁺ (48), 43(20), 41(25).

Epoxidation of 1 (70 mg) with *m*-chloroperoxybenzoic acid (40 mg) in CH₂Cl₂ (10 ml) at room temperature for 2 h afforded, after usual work up and purification by chromatography, an epoxide identical in all aspects to natural 14 (42 mg).

Acetate **15**.—Compound **14** (127 mg) in Ac₂O/Py as above gave **15** (131 mg); mp 173-174° (hexane); $[\alpha]D - 25^{\circ}$ (CHCl₃, c 1.0); ir ν max (KBr) 2920, 2860, 1730, 1710, 1650, 1450, 1360, 1240, 1160, 1040, 980, 840 cm⁻¹; ¹H nmr (60 MHz) δ 0.86 (6H, s, H-18 and H-19), 1.10 (3H, s, H-20), 1.93 (3H, br s, H-5'), 2.03 (3H, br d, J=7 Hz, H-4'), 2.03 (3H, s, MeCO), 2.79 (2H, s, H-17), 4.46 (1H, dd, J=9 and 7 Hz, H-3), 5.02 (1H, br s, H-15), 6.13 (1H, br q, J=7 Hz, H-3'); ms m/z (%) 444 [M]⁺ (2), 361 [M-Ang]⁺ (2), 344 [M-AngOH]⁺ (12), 329 [344-Me]⁺ (7), 326 [344-H₂O]⁺ (5), 301 [361-AcOH]⁺ (9), 284 [344-AcOH]⁺ (10), 269(12), 251(10), 135(13), 83 [Ang]⁺ (100), 55 [Ang-CO]⁺ (22), 43(10).

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